

## ENZYMATIC AND BIOLOGICAL ACTIVITY ANALYSIS OF ENDOPHYTIC FUNGI ISOLATED FROM PIPER NIGRUM

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### ABSTRACT

Piper nigrum is well known for its metabolite richness and medicinal property. So the endophytic fungi associated with the plant can also be expected to have promising bioactive metabolites. In this part of the study, endophytic fungi isolated from P. nigrum were screened for the antimicrobial properties against Staphylococcus aureus, Bacillus subtilis and Vibrio cholerae. For this, the culture extracts were prepared made from the fungal isolates Colletotrichum sp., Diaporthe sp., Phomopsis sp., Mycosphaerella sp., Fusarium sp., Pleosporales sp. and Pseudocercospora sp. The isolates Fusarium sp. and Phomopsis sp. were found to have highest activity against the pathogens screened. However, 80% of isolates were found to be positive for the production of asparaginase. 40% were positive for amylase and protease and 20% showed cellulase and laccase production.

**Keywords:** Endophytic fungi, Piper nigrum, pathogens, antimicrobial activity

### 1. INTRODUCTION

Bioactive natural products produced by endophytic fungi can have tremendous applications. Endophytic fungi isolated from medicinal plants have been recognized as beneficial source of bioactive metabolites (Wen et al., 2022). Recent investigations have proved endophytic microorganisms as potential sources for human beneficial compounds including antibiotics, anti malaria and anti carcinogens (Strobel, 2003). Hence endophytic fungi from medicinal plants have been widely studied for novel secondary metabolites. The emerging information from endophytic studies indicate the chemical nature of bioactive compounds as alkaloids, steroids, terpenoids, isocoumarins, quinones, flavonoids, phenylpropanoids, peptides, phenolics, and aliphatics (Hashem et al., 2023).

Endophytic fungi have the ability to produce valuable enzymes also. Enzymes derived from fungi are used in food, beverages, confectionaries, textiles and leather industries to simplify the processing of raw materials. They are often more stable than enzymes derived from other sources (Toppo et al., 2024). Enzymes such as cellulase, lipase and chitinase are common enzymes produced by endophytic Beauveria, Lecanicillium, Metarhizium and Trichoderma, isolated from various plant species (Bhadra et al., 2023). Asparaginase and tyrosinase are two valuable enzymes with increasing demand due to their wide use in medicine (Juluri et al., 2023). Other major hydrolytic enzymes like amylase, cellulase and laccase of endophytic origin also have various industrial applications (Hawar et al., 2022).

The endophytes from medicinal plants have become an important topic for metabolite discovery (Gouda et al., 2016). Black pepper (Piper nigrum), which belongs to the family Piperaceae is known to have traditional use for the treatment of malaria, gastro intestinal disorders, epilepsy and asthma (Takooree et al, 2019). So endophytic fungi present in this plant can expect to have remarkable biosynthetic potential. In this part of the study, anti microbial, enzyme activity and plant growth promoting features of endophytic fungi isolated from the Piper nigrum was carried out.

### 2. MATERIALS AND METHODS

#### 2.1 Fungi selected for the study

Among 18 endophytic fungal isolates, seven isolates like Colletotrichum sp.(KC213984), Diaporthe sp. (KC213981), Phomopsis sp. (KC213980), Mycosphaerella sp.(KC213982), Fusarium sp. (KC213986), Pleosporales sp. (KC213985) and Pseudocercospora sp. (KC213983) were selected for the study.

#### 2.2 Determination of Enzyme activity

Extra cellular enzyme production property was screened for selected endophytic fungal species from Piper nigrum. The isolates were screened for cellulase, laccase, lipase, protease, amylase and asparaginase activity (Hawar et al., 2022).

##### 2.2.1 Amylase

The amylase activity was screened by inoculating the selected isolates on GYP agar medium (1g/L glucose, 0.1g/L yeast extract, 0.5g/L peptone, 16g/L agar, 1000 mL distilled water, pH 6) with 2% soluble starch (Sunitha et al.,

2012). After 3-5 days incubation time, the fully formed cultures were flooded with 1% iodine in 2% potassium iodide. Formation of clear halos was considered as positive results.

### 2.2.2 Cellulase

The cellulase activity was determined by inoculating the isolates on Czapek dox medium (30g/L sucrose, 3 g/L NaNO<sub>3</sub>, 1 g/L K<sub>2</sub>HPO<sub>4</sub>, 0.5 g/L MgSO<sub>4</sub>, 0.5 g/L KCl, 0.01 g/L FeSO<sub>4</sub>, 16 g/L Agar) amended with 0.5% Na-carboxy methyl cellulose (Mrudala et al., 2011). The plates after incubation were flooded with 0.1% congo red and destained with 1M sodium chloride. Formation of clear zone was considered as positive results.

### 2.2.3 Laccase

For Laccase activity, the isolates were grown on GYP agar medium (1 g/L glucose, 0.1 g/L yeast extract, 0.5 g/L peptone, 16 g/L agar, pH 6) and 1-naphthol, 0.005% is added along with the media (Jayaram et al., 2023). The oxidation of 1-naphthol by laccase enzyme produced by the isolates was observed.

### 2.2.4 Lipase

The lipase activity was analyzed by growing the selected isolates on the peptone agar media (10 g/L peptone, 5 g/L NaCl, 0.1 g/L CaCl<sub>2</sub>. 7H<sub>2</sub>O, 16 g/L agar, pH 6) as described by Mobarak-Qamsari et al., (2011). To the sterilized peptone agar culture media, previously sterilized Tween 20 was added a final concentration of 1% (v/v). The media was inoculated with the isolates and incubated. The formation of halos around the colony was considered as positive results.

### 2.2.5 Protease

The proteolytic activity was observed by growing the isolates on GYP agar media (1g glucose, 0.1 g/L yeast extract, 0.5 g/L peptone, 16 g/L agar, pH 6) supplied with 0.4% gelatin. To the sterilized culture media, separately sterilized 8g of gelatin in 100 mL distilled water was added (Sunitha et al., 2013). After incubation, the culture was flooded with saturated aqueous ammonium sulphate. Formation of clear zone around the colony was then observed as indication of hydrolysis of gelatin in media.

### 2.2.6 Chitinase

For chitinase activity, the endophytic fungi were grown on colloidal chitin agar medium (2g/L colloidal chitin, 16 g/L agar) (Narayana et al., 2009). Formation of clear zone surrounding the colony after incubation was considered positive for chitinase activity.

### 2.2.7 Asparaginase

For this, the selected endophytic fungal isolates were cultured on PDA for 7 days. The 5 mm disc of mycelium was transferred to Modified Czapek Dox's (MCD) agar (2.0 g/L glucose, 10.0 g/L L-asparagine, 1.52 g/L KH<sub>2</sub>PO<sub>4</sub>, 0.52 g/L KCl, 0.52 g/L MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.001 g/L CuNO<sub>3</sub>. 3H<sub>2</sub>O, 0.001 g/L ZnSO<sub>4</sub>.7H<sub>2</sub>O, 0.001 g/L FeSO<sub>4</sub>.7H<sub>2</sub>O, 16g/L Agar) supplemented with phenol red (0.009% final concentration) indicator (Juluri et al., 2022). Control plates were MCD agar without asparagine. All the plates were incubated at 30°C. Formation of pink zone radius was considered as positive isolates.

## 2.3 Antibacterial Activity

The crude extracts of endophytic fungi prepared as explained above were tested against *Bacillus subtilis*, *Staphylococcus aureus*, and *Vibrio cholerae*. Anti microbial activity was determined by using well diffusion method. The overnight cultures of test organisms were inoculated in to Muller Hinton Agar (2.00 g/L beef extract, 17.50 g/L acid hydrolysate of casein, 1.50 g/L starch, 12g/L agar) plates using sterile cotton swab. About 6 mm size wells were made and 30µL of crude extract was added into it and kept for incubation at 37°C for 24 h. Same volume of methanol alone was used as control. Anti microbial activity was analyzed based on the zone of inhibition formed (Balouiri et al., 2016).

## 3. RESULTS AND DISCUSSION

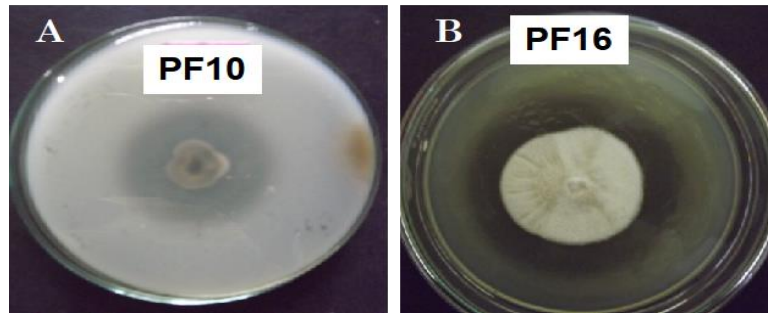
In this part of the study, seven endophytic fungi were screened for the presence of extracellular enzymes amylase, cellulase, laccase, chitinase, asparaginase and protease by culturing on specific media (**Table 1**).

Two endophytic fungal strains (*Pleosporales* sp. and *Mycosphaerella* sp.) were identified to have protease activity (**Fig. 1**). *Mucor* sp. was previously reported to be the best producer of protease, and with previous reports 82% of endophytic fungi may likely have proteolytic activity. This has application in waste treatment, detergents, cosmetics and leather manufacture, the pharmaceutical and food industries (Orlandelli et al., 2015). Among the seven endophytic fungi, *Phomopsis* sp. and *Fusarium* sp. showed promising amylase activity (**Fig. 2**) while only *Pseudocercospora* sp. was found to have cellulase and laccase activity. Amylase is a major enzyme used in starch processing industries for

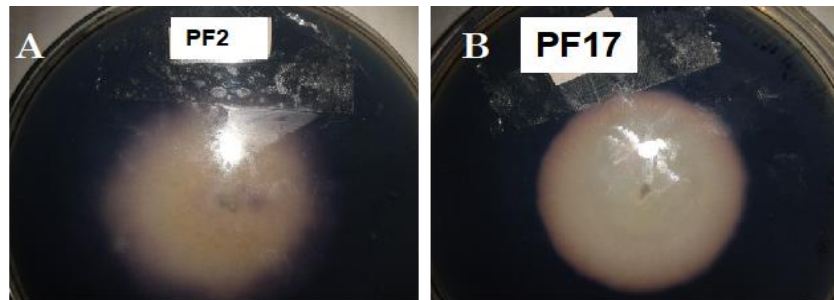
the hydrolysis of polysaccharides into simple sugars (Sunitha et al., 2012). The cellulases produced by many terrestrial fungi are being used in paper industry (Rajput et al., 2016).

Asparaginase is used as a chemotherapeutic agent against acute lymphoblastic leukemia and lympho-sarcoma. The enzyme catalyzes the hydrolysis of L-asparagine into L-aspartic acid and ammonia (Juluri et al., 2022). Here, the enzyme production was observed for five endophytic fungal species like *Pseudocercospora* sp., *Phomopsis* sp., *Mycosphaerella* sp., *Fusarium* sp. and *Pleosporales* sp. (Fig. 3).

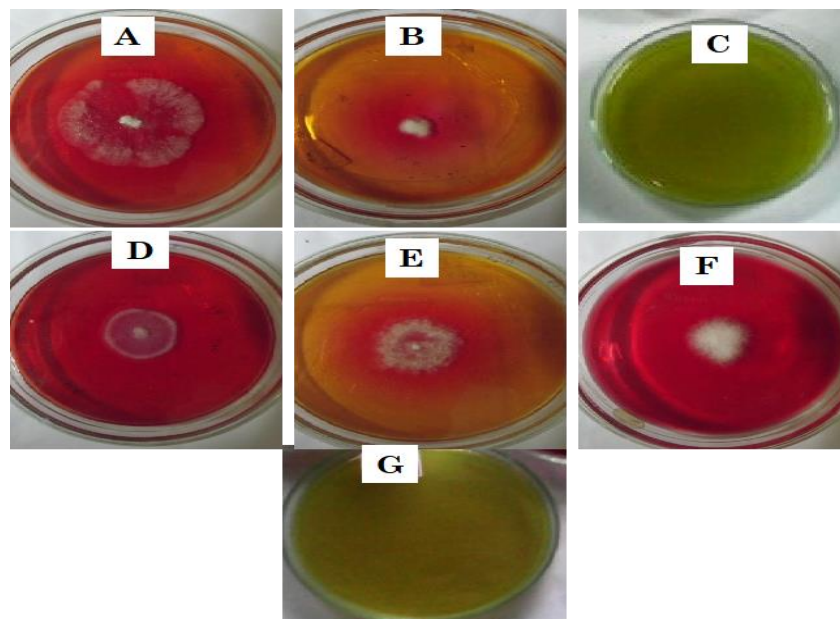
Fungal enzymes are gaining importance in agriculture, industry and also for medical applications as because of their stability when compared to enzymes plants and animals. Fungal enzymes are used in manufacturing of food, beverages, confectioneries and textile. Hence screening of endophytic fungi for enzymes, antibiotics, anti- cancer drugs will have significant application.



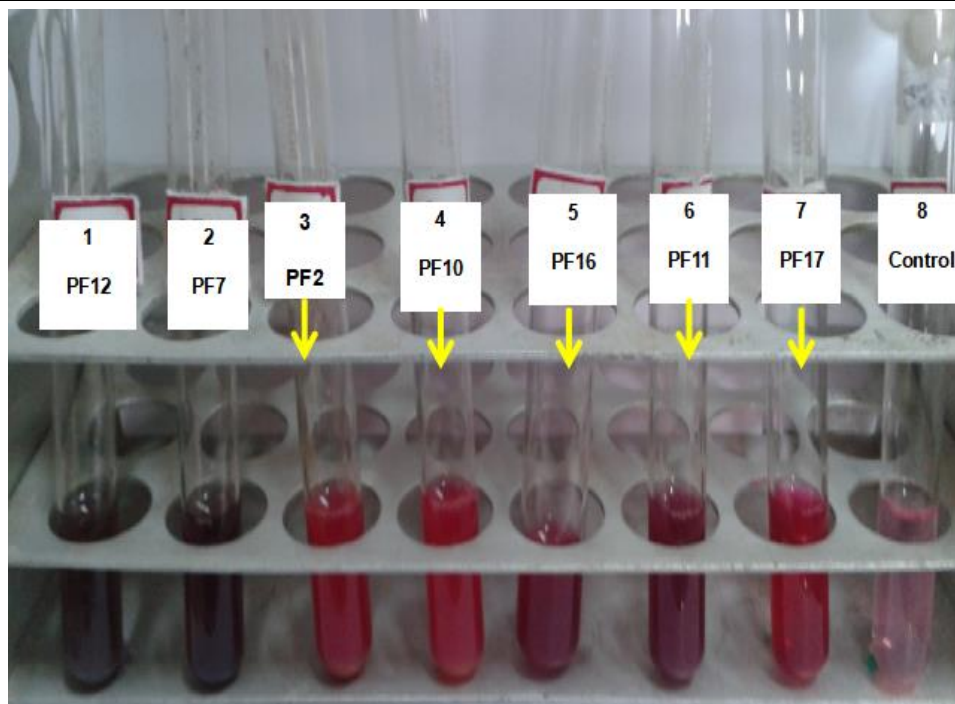
**Fig. 1:** Protease activity of endophytic fungi isolated from *Piper nigrum*. A – *Mycosphaerella* sp. (PF 10); B – *Pleosporales* sp. (PF 16).



**Fig. 2:** Amylase activity of endophytic fungi isolated from *Piper nigrum*. A – *Phomopsis* sp. (PF 2); B – *Fusarium* sp. (PF 17).



**Fig. 3:** Endophytic fungal isolates with L-asparaginase production. A - *Phomopsis* sp. (PF 2) +ve ; B – *Pseudocercospora* sp.(PF11) +ve; C-*Colletotrichum* sp. (PF12 no activity); D- *Mycosphaerella* sp. (PF10) +ve ; E - *Pleosporales* sp. (PF16) +ve; F-*Fusarium* sp.(PF17) +ve ; G- *Diaporthe* sp. (PF7 no activity).



**Fig. 4:** Qualitative analysis on Asparaginase enzyme production by selected endophytic fungi; dark pink indicates the presence of asparaginase 1- Colletotrichum sp. (PF12 no activity); 2-Diaporthe sp. (PF7 no activity); 3- Phomopsis sp. (PF 2) +ve; 4) Mycosphaerella sp. (PF10) +ve; 5- Pleosporales sp. (PF16) +ve; 6-Pseudocercospora sp. (PF11) +ve ; 7- Fusarium sp. (PF17) +ve; 8- Control

**Table 1:** Summary of enzyme activity of selected endophytic fungi

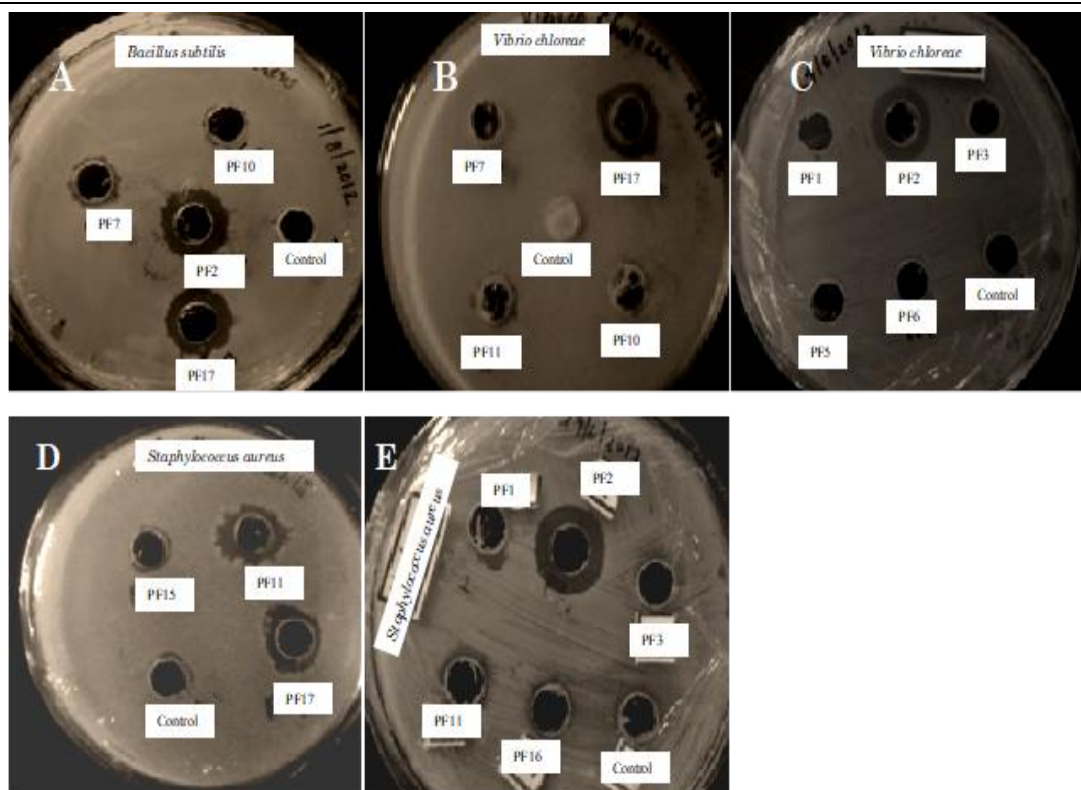
Isolates	Enzyme activity screened				
	Amylase	Protease	Laccase	Cellulase	Asparaginase
PF2 (Phomopsis sp.)	++	-	-	-	++
PF10 (Mycosphaerella sp.)	-	++	-	-	++
PF11 (Pseudocercospora sp.)	-	-	+	+	++
PF12 (Colletotrichum sp.)	-	-	-	-	-
PF7 (Diaporthe sp.)	-	-	-	-	-
PF16 (Pleosporales sp.)		++	-	-	+
PF17 (Fusarium sp.)	++	-	-	-	++

Enzyme activity of selected endophytic fungi from Piper nigrum. +indicates small activity; ++ indicates good activity; - indicates no activity;

Screening for antibacterial activity of the endophytic isolates was conducted by agar well diffusion method against both gram positive and negative bacteria. The crude extracts of Phomopsis sp., Mycosphaerella sp., Colletotrichum sp., Diaporthe sp., Pseudocercospora sp., Fusarium sp. and Pleosporales sp. showed varying levels of antibacterial activity towards the test organisms like Bacillus subtilis, Staphylococcus aureus, and Vibrio cholerae (**Table 2**). Most of the fungal extracts were active against Staphylococcus aureus and Bacillus subtilis. However among the isolated strains crude extract of Phomopsis sp. and Fusarium sp. were active against Staphylococcus aureus, Vibrio cholerae and Bacillus subtilis with 30-32 mm of zone of inhibition (**Fig. 5**). Pleosporales sp., Pseudocercospora sp. and Colletotrichum sp. have highest activity against Bacillus subtilis zone of inhibition about 20-25 mm and Diaporthe have activity against Staphylococcus aureus also. The endophytic Colletotrichum sp. was found to have activity against phytopathogen Rhizoctonia sp.

Jalil et al., (2022) have reported the antimicrobial compounds produced by endophytic fungi are extractable with ethyl acetate. The endophytes such as P. citrinum and N. dimidiatum have been reported to produce compounds active against the plant pathogenic fungi T. cucumeris and G. zeae (Xia et al., 2022).





**Fig. 5:** Antimicrobial activity of endophytic fungi isolated from *P. nigrum* A) *Phomopsis* sp. (PF2) and *Fusarium* sp. (PF17) against *Bacillus subtilis* B) *Fusarium* sp. (PF17) against *Vibrio cholerae* C) *Phomopsis* sp. (PF2) against *Vibrio cholerae* D) *Fusarium* sp. (PF17) active against *Staphylococcus* sp. E) *Phomopsis* sp. (PF2) against *Staphylococcus* sp.

**Table 2:** Summary of the antibacterial activity exhibited by endophytic fungal isolates from *P. nigrum*

Name of organism	Antibacterial activity analysis			Antifungal activity
	<i>Staphylococcus aureus</i>	<i>Vibrio cholerae</i>	<i>Bacillus Subtilis</i>	<i>Rhizoctonia</i> sp.
<b><i>Phomopsis</i> sp. (PF2)</b>	++	++	+++	-
<b><i>Mycosphaerella</i> sp. (PF10)</b>	-	-	-	-
<b><i>Pseudocercospora</i> sp. (PF11)</b>	-	-	++	-
<b><i>Fusarium</i> sp. (PF17)</b>	++	++	+	-
<b><i>Diaporthe</i> sp. (PF7)</b>	++	-	-	-
<b><i>Colletotrichum</i> sp. (PF12)</b>	-	-	++	+++
<b><i>Pleosporales</i> sp. (PF16)</b>	+	-	+++	-

Antimicrobial activity of selected endophytic fungi from *Piper nigrum*. + indicates > small inhibition; ++ indicates moderate inhibition; +++ indicates good inhibition; – indicates no activity.

The crude extract of endophytic fungi was found to have activity against *Bacillus subtilis*, *Staphylococcus aureus* and *Vibrio cholerae*. Among this, highest inhibition was observed to be against *Staphylococcus aureus* and *Bacillus subtilis*. Many endophytic fungi have the ability to produce metabolites with antimicrobial activity (Jha et al., 2023). Endophytic fungi from *Terminalia brownie* showed significant activity against *S. aureus*, *E. faecalis*, *P. aeruginosa*, *E. coli* and *C. albicans* (Kouipou et al., 2019).

These reports are supportive to the importance of endophytic fungi from medicinal plants as promising source of anti microbial agents. Guimaraes et al., (2008) have reported the endophytic fungi isolated from *Viguiera arenaria* and *Tithonia diversifolia* to have activity against *S. aureus* and *E. coli*.

#### 4. CONCLUSION

Among 18 endophytic fungi from *P. nigrum*, *Colletotrichum* sp. *Diaporthe* sp. *Phomopsis* sp. *Mycosphaerella* sp. *Fusarium* sp. *Pleosporales* sp. and *Pseudocercospora* sp. were selected for further studies. Screening for anti microbial studies revealed *Phomopsis* sp. and *Fusarium* sp. to have activity against three bacterial pathogens such as *S. aureus*, *V. cholera* and *B. subtilis*. The endophytic fungi were identified to have the ability to produce enzymes screened. The remarkable production of asparaginase observed in the study is very significant as it is having anticancer effect.

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